RESEARCH COMMUNICATION

Effects of Yiqi Chutan Tang on the Proteome in LEWIS Lung Cancer in Mice

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Abstract

In order to verify effects of Yiqi Chutan Tang on lung cancer and assess molecular mechanisms involved we focused on size, tumor weight and the numbers of lung metastases and differential expression protein spot information acquired by two-way fluorescence with a tumor difference gel electrophoresis (2D-DIGE) system, and differentially expressed proteins were identified by matrix-assisted laser desorption ionization time of flight mass spectrometry (MALDI-TOF-TOF). Differences were finally verified by Western blot and fluorescence quantitative PCR. We found that tumor size, tumor weight in Yiqi Chutan Tang treatment group were significantly less than that in model group (p<0.01), with a tumor growth inhibition rate of 57.2% . For gel diagram analysis of 2D-DIGE system, compared with model group, there were 44 expressed differentially protein spots, of which 6 were up-regulated and 38 were down-regulated. Among these proteins, 37 (30 down-regulated and 7 up-regulated) were successfully identified by MALDI-TOF-TOF. In conclusion, Yiqi Chutan Tang effects on LEWIS lung cancer appeared highly related to down-regulated expression of Hspd1, prolyl 4-hydroxylase, protein disulfide-isomerase A3 precursor, EG433182, heat shock protein 5 precursor, heat shock protein 9 and stress-induced phosphoprotein 1.

Keywords: Yiqi Chutan Tang - proteome - lung cancer - anti-tumor effects

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Introduction

Clinical studies have shown that combined treatment of Chinese medicine with surgery and chemotherapy could significantly improve the survival rate and prolong survival of patients with lung cancer (Zhou and Lin, 2008; Zhang and Yu, 2010).

Yiqi Chutan Tang (YQCT) is composed of Codonopsis, raw Pinellia, Honeycomb, and so on. The varieties of all materials in prescription used in the experiment were selected according to “Chinese Pharmacopoeia” 2005 Edition, and the YQCT was also identified by the experts. It has already been verified that Codonopsis which is major component of Yiqi Chutan Tang had better effects in the treatment of lung cancer (Li et al., 2008). In order to verify the effects of Yiqi Chutan Tang on lung cancer and the molecular mechanism involved, we observe the anticancer effect of Yiqi Chutan Tang on Lewis lung cancer. Few research has been conducted on the antitumor effects of Yiqi Chutan Tang in vivo, especially the underlying mechanism of which some molecules participating in the anti-tumor effect. So the objectives of this study were identify the differentially expressed proteins using 2D-DIGE system and MALDI-TOF-TOF (Suckau et al., 2003; Yang et al., 2007; Kondo and Hirohashi, 2009), further analysis the mechanism which Yiqi Chutan Tang exert its effect in the process.

Materials and Methods

Substances

Yiqi Chutan Tang (YQCT) ingredients were boiled three times with purified water, 30min each time, and the amount of purified water supplied respectively was 10 times, 8 times, 8 times than the amount of the herbs. After boiling liquid collection, filtration, it was finally concentrated to crude drug with the amount of 2g/ml, reserved at 4 °C for use.

Cell line

Lewis lung cancer cells (the Cancer Center of West China Medical University, China) were incubated with RPMI-1640 (Gibco, USA) which contained 10% FBS. Cells were incubated in a CO2 Water Jacketed Incubator (Forma Scientific) and grown at 37°C in an O2 5%/CO295% air atmosphere. Cell viability was determined by trypan blue exclusion analysis.

Animals

The animals were bred and maintained in a HEPA filtered environment with cages, and received human care according to the criteria outlined in the “Guide for the Care and Use of Laboratory Animals” prepared by the National Academy of Sciences and published by the National Institute of Health.

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Establishment of the lung cancer model

A total of 40 SPF C57BL mice (SCXK (Chongqing) 2007 – 0001), including evenly male and female, 18 ~ 20 g, were feeding adaptively for 1 week. A single dose of 1×10⁷ Lewis lung cancer cells were inoculated subcutaneously into the right axillary of C57BL mice. Then Yiqi Chutan Tang (3.0 g·kg⁻¹·d⁻¹, treatment group) and saline (model group) were given daily by gavage for 21 days.

Tumor growth and metastasis inspection

We performed the experiment as previously (Berger et al., 2010). Briefly, during the period of the study, body weight of the mice will be measured every three days, the amount of drug was adjusted according to the body weight of mice. After mice were sacrificed, the longest diameter (a) and the shortest diameter (b) was measured with a vernier caliper, the tumor volume was calculated according to formula: V=0.5 ab², and the inhibition rate of tumor growth was also calculated according to formula: tumor growth inhibition rate (%) = (average tumor weight of normal saline group - average tumor weight of treatment group) / the average tumor weight of normal saline group × 100%. At the meanwhile, the number of bilateral lung cancer metastasis were counted under 100 × microscope. The number of bilateral lung cancer metastasis in each field were counted in a total of 5 high-power fields (magnification ×100) per region under ocular micrometers (Olympus).

Preparation of proteins

The protein mixture of same quantity was extracted from tumor tissues of treatment group and model group with 1000ul lysis buffer (8M Urea, 4% CHAPS, 40mM tris PH8.5, 0.5% carrier ampholyte (3-10NL)). The protein concentration of samples was determined with the Bradford protein assay (Bio-Rad, USA), using bovine gamma globulin as the standard. The CyDye DIGE Fluor minimal dye (Amersham, USA) was reconstituted in 10 μl of DMF by centrifuging at 12,000g for 30 s, a deep color of the dye at this point was ensured (Cy2–yellow, Cy3–red and Cy5–blue).

Two-dimensional electrophoresis isoelectric focusing (IEF) was carried out using commercially available, dedicated apparatuses: IPGphor Protean IEF Cell (Amersham, USA). IPG strips (Amersham, USA) were used according to manufacturer instructions. About 200 μg of sample for gel were applied to immobilized pH 3 to 10 nonlinear by overnight re-hydration at 50 V. With Protean IEF Cell, focusing was done initially at 500 V for 1 hour, then the voltage was increased quickly to 1 000 V for 1 hour, 8 000 V for 1 hour, 65 000 V for 1 hour with a total of 35 kVh. After the first-dimensional IEF, IPG gel strips were transferred to the equilibration solution containing 2.5% iodoacetamide and then were placed on 12% polyacrylamide gel electrophoresis (PAGE) and transferred to PVDF membrane (Millpore, USA). The gel was stained with coomassie blue to document equal protein loading. Membrane was blocked with 5% dry milk and 0.1% Tween 20 (USB,USA) in PBS and incubated with rabbit anti-rat HSP-60 Ab, P4HB Ab, PDIA3 Ab or rabbit anti-rat β-actin Ab (Santa Cruz, USA). The filters was developed until the bromophenol blue dye marker had reached the bottom of the gel.

Protein visualization and image analysis

Protein patterns in the gels were recorded as digitalized images using a high-resolution scanner (Typhoon, Amersham, USA). Gel image matching was done with Decyder 2D software (Amersham, USA). Scanned gel images were processed to remove backgrounds, staining on the gel borders and to automatically detect spots. For all spot intensity calculations, normalized values were used. Normalization of spot intensity was done with Loess Regression Method and normalized spot intensities were expressed in ppm.

MS Sample Preparation and Mass Spectrometry Analysis

In-gel digestion of protein spots on Coomassie gels was carried out with 160ng of Porcine Modified Trypsin (Sigma) in 10% ACN and 25mM NH4HCO3. All mass spectra were acquired on an Applied Biosystems 4700 Proteomics Analyzer equipped with TOF/TOF ion optics and a diode pumped Nd:YAG laser with 200 Hz repetition rate. Peptides were identified using ProteinScape™ database (Protagen, Germany) and Mascot search engine to cross-validate or consolidate the identification results through the complementary use of several software packages.

Western blot analysis

Protein was extracted from tumor tissues with PBS TDS buffer (50 mM Tris, 150 mM NaCl, 0.1% SDS, 1% sodium deoxycholate, and 1% triton X-100, PH 7.2). Protein (30μg/ sample) in SDS-loading buffer (50 mM Tris, PH 7.6, 10% glycerol, 1% SDS) were subjected to 12% SDS-polyacrylamide gel electrophoresis (PAGE) and transferred to PVDF membrane (Millpore, USA). The gel was stained with coomassie blue to document equal protein loading. Membrane was blocked with 5% dry milk and 0.1% Tween 20 (USB,USA) in PBS and incubated with rabbit anti-rat HSP-60 Ab, P4HB Ab , PDIA3 Ab or rabbit anti-rat β-actin Ab (Santa Cruz, USA). The filters
were washed and incubated with horseradish peroxidase goat anti-rabbit Ab (Amersham, USA).

Quantitative RT-PCR
Total tissue RNA was prepared using TRIzol reagent (Shine Gene, USA). Five μg of RNA was reverse-transcribed into cDNA using oligo (dT) primers with Omniscript™ reverse transcriptase (Shine Gene, USA). Quantitative-PCR was performed using the thermocycler (Shine Gene, USA). In a final reaction volume of 25μl, the followings were added (Shine Gene, USA): Taq polymerase, 1×Taq buffer, 125 MmNTP, SYBR Green I (Molecular Probes), and Fluorescein, together with cDNA and primers. Amplification conditions were: 94˚C (4min), 35 cycles of 94˚C (20s), 60˚C (30s), 72˚C (30s). The primers used to amplify a specific 100-300bp fragment of EG433182, HSP5P, HSP9, STIP1 and β-Actin are the following: EG433182 5', agtccttcgtccagaactaccc, 3', gccttggcaatccgctta; HSP5P 5', aggtgggcaaaccaagacat, 3', agtccagcaatagtgccagc; HSP9 5', aatgattcacagcgacaggc, 3', cctcctaagaaagtgtccccat; STIP1 5', etgagcctcacaagacag, 3', cactcggggttcggtgct; β-Actin 5', tagacctacaccaagacag, 3', atgtcacgcacgatttccc.

Statistical analysis
All values are expressed as mean ± SD. Data were analyzed with an unpaired two-tailed Student’s t test. P<0.05 was considered to be statistically significant

Results
For Yiqi Chutan Tang treatment group, tumor size (1.21±0.43 vs 2.83±0.35 cm3), tumor weight (0.89±0.31 vs 2.08±0.28 g) and the numbers of lung metastases (5.30±2.06 vs 8.60±1.84) were significantly less than that in model group (p <0.01), and the tumor growth inhibition rate was 57.2%.

Two-dimensional electrophoresis of proteins
The 2-DE proteomics carries the advantage of visualizing changes in Mw and pI of a protein, which we find helpful in highlighting biologically significant processes. This electrophoresis technique has been applied successfully to identify oncoproteins in serum and tissues. In order to eliminate individual differences, we mixed respectively the samples of treatment group and model group. By gel diagram analysis of 2D-DIGE system, as a master gel, G1 editing a total of 1139 points, and G2 editing a total of 1186 points, 921 points were matched. Then we further compared treatment group with model group, there were 44 expressed differentially protein spots, of which 6 protein spots were up-regulated and 38 protein spots were down-regulated in the treatment group (Figure 1).

MALDI-TOF-TOF analysis of proteins
The mass list from the mass spectra (MALDI-TOF) was used for peptide mass fingerprinting for protein identification (see Table 1). Some of the precursor ions from MS were further processed by MS/MS (MALDI-TOF-TOF) to generate confirmatory sequence information. Striking differences in the MALDI MS spectra of peptides were observed between treatment group and model group. Among the proteins with a total of 37, 30 proteins expression are down-regulated, including vimentin, prolyl 4-hydroxylase, alpha-globin, tubulin beta 2, T-cell receptor alpha chain precursor V region (4.C3), protein disulfide-isomerase A3 precursor, phosphoglycerate kinase, heterogeneous nuclear ribonucleoprotein H1, EG433182 protein, mKIAA4049 protein stress-induced phosphoprotein 1, heat shock protein 5 precursor, phosphoglucomutase 5, heat shock protein 9, mCG116950, phosphoglycerate kinase 1. PREDICTED: hypothetical protein LOC73530, Hspd1 protein, and so on; 7 proteins expression were up-regulated, including Rho GDP dissociation inhibitor (GDI) alpha, alpha-1-globin, biliverdin reductase B, Parkinson disease (autosomal recessive, early onset) 7, hypothetical protein LOC433182, Shc SH2-domain binding protein 1, isoform CRA_a.

Western blot analysis
Of those expressed differentially proteins, we focused on the drastically changed proteins and compared their expression between treatment group and model group by Western blots. It was obvious that the expression of Hspd1, prolyl 4-hydroxylase and protein disulfide-isomerase A3 precursor decreased after treatment (Figure 2).

table1

| Protein name | Peptide mass | R. | Source | up-regulated | down-regulated | Full length
|--------------|--------------|---|--------|-------------|----------------|-------------
| EG433182     |              |   |        |             |                |             
| HSP5P        |              |   |        |             |                |             
| HSP9         |              |   |        |             |                |             
| STIP1        |              |   |        |             |                |             
| β-Actin      |              |   |        |             |                |             
| Rho GDP dissociation inhibitor (GDI) alpha | | | | | | 
| α-1-globin | | | | | | 
| Biliverdin reductase B | | | | | | 
| Parkinson disease (autosomal recessive, early onset) 7 | | | | | | 
| Hypothetical protein LOC433182 | | | | | | 
| Shc SH2-domain binding protein 1, isoform CRA_a | | | | | | 

qRT-PCR gene expression detection in two groups
Gene expression detection is an alternative method
Discussion

In this study, we clearly observed the inhibitory effect of Yiqi Chutan Tang on Lewis lung cancer in C57 BL mice, tumor size, tumor weight and the numbers of lung metastases were decreased significantly. On consideration of clinical need, it is more important to study the anti-tumor effects of compound Chinese herbal medicine rather than a single herb (Ji et al., 2009).

For further analysis, we detected a very interesting phenomenon that several kinds of HSPs including heat shock protein 60, heat shock protein 5 precursor and heat shock protein 9, involving in the Lewis lung cancer tumorigenesis. At the meanwhile, we also found the expression of Stress-induced-phosphoprotein (STIP1), also called Hsp90/Hsp70-organizing protein (Hop) changed drastically in our research. We focus on the role of HSPs and related molecules in tumorigenesis in many different ways.

A first involvement of hsp in T cell-mediated immunity was demonstrated by the pioneering studies of Srivastava and colleagues, who showed that many HSPs allows them to deliver tumour antigens very effectively to antigen presenting cells. In murine systems, vaccination with heat shock proteins (HSPs) such as glucose-regulated protein (GP)96, HSP70, and HSP90 from cancer tissues but not from normal tissues induces specific immunity and CTL activation (Li, 1997; Belli et al., 2002). The specificity of the induced CTLs relies on the peptides chaperoned by these HSPs. This property allows CTL activation without support. The length of encoded protein is conserved, open reading frame as well as strong transcriptional support. The length of encoded protein is conserved, open reading frame as well as strong transcriptional support. The length of encoded protein is conserved, open reading frame as well as strong transcriptional support. The length of encoded protein is conserved, open reading frame as well as strong transcriptional support. The length of encoded protein is conserved, open reading frame as well as strong transcriptional support.

Hypoxia/ischemia is a common feature of solid tumors. The cellular response to hypoxic stress is controlled by a family of prolyl hydroxylases (PHD) and the transcription factor hypoxia-inducible factor 1 (HIF1). The oxygen requirement suggests that PHDs are the cellular “sensors” for hypoxia(Gaiser et al., 2009) while HIF1 regulates a balance between cellular adaptation, through modulation of various pathways. Moreover, Prolyl 4-hydroxylase (P4H), as enzymic catalysts of prolyl hydroxylation, mediated an irreversible reaction that is the most common posttranslational modification in humans (Baksh et al.,1995; Chang et al., 2007). The biological consequences of prolyl hydroxylation vary widely, and include altering protein conformation and protein-protein interactions, and enabling further modification. In addition, an increase of hsp9 mRNA in the transcript level is also reported in response to glucose deprivation.
In summary, in this study we successfully completed the purpose of screening differentially expressed proteins, moreover, we identified those inflammation-induced proteins that have abnormal changes in 2D-DIGE analysis. In order to clarify the role of these proteins in pathogenesis and to estimate whether these proteins are useful for developing new diagnostic markers or therapies, further study is needed.

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